



Demi-embryo reconstitution, a factor to consider for the success of embryo bisection. Review



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Abstract:

For many years it has been sought to increase the reproductive efficiency of livestock using biotechnologies such as embryo bisection. However, despite its potential in livestock, its level of adoption is limited. The present work reviews the importance of demi-embryo reconstitution, after bisection, and the main factors that limit its success in livestock. It is possible to increase its level of adoption if it is possible to increase the efficiency currently obtained with this technique, this can be achieved by making a more precise selection of the embryos subjected to bisection. Embryo quality is one of the most important factors related to the potential to reconstitute into viable demi-embryos after bisection, which can be used with greater reliability in embryo transfer programs.

Key words: Embryo bisection, Demi-embryo reconstitution, Embryo development.

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Introduction

Embryo bisection is a reproductive biotechnology that allows obtaining identical demi-embryos, to be used in research⁽¹⁾ or the livestock industry⁽²⁾. The aim of embryo bisection is to increase the number of semi-embryos available for transfer and therefore, the offspring of genetically superior animals^(1,3,4). This technique can be applied to embryos developed in the morula or blastocyst stage and consists of obtaining two similar halves by mechanical bisection^(5,6,7). Morula bisection can be carried out in any position of the embryo, due to its symmetric morphology⁽⁸⁾. In the case of blastocysts is important the symmetrical orientation of the embryo to obtain a proportional distribution of the inner cell mass (ICM) and trophoctoderm (TE) in the resulting demi-embryos⁽⁹⁾.

Embryo bisection is performed in embryos of several species^(10,11,12), in order to increase embryo availability⁽¹²⁾, pregnancy rate⁽¹³⁾, and the number of offspring^(5,14,15). However, there are studies where the survival of demi-embryos was low^(16,17), even lower compared to the use of whole embryos⁽¹⁸⁾. This could be associated with the fact that embryo bisection is an invasive technique⁽⁶⁾ and the procedure causes cellular damage⁽¹⁹⁾. Therefore, the success of the technique could be influenced by factors associated with the original embryo^(20,21,22) and its ability to reconstitute itself in the resulting demi-embryos⁽²³⁾. The objective of this review is to highlight the importance of demi-embryo reconstitution in embryo bisection programs, as well as to discuss the main factors that influence its success.

Importance of embryo bisection in livestock

Embryo bisection has been carried out in different livestock species of interest, such as rabbits⁽¹⁹⁾, sheep⁽²⁴⁾, bovines⁽¹²⁾, goats⁽¹⁰⁾, equines⁽²⁵⁾, swine⁽²⁶⁾, and even in humans⁽²²⁾. Despite it is an invasive technique, it is practical and it does not require cell reprogramming like cloning⁽⁶⁾. Embryo bisection in livestock allows to produce identical twins for experimental use⁽¹⁴⁾, reducing the number of animals needed per treatment for comparison tests⁽²⁷⁾ or to increase the availability of transferred embryos⁽²⁸⁾. In addition, obtaining identical twins facilitates the evaluation of sires or maternal trait tests⁽²⁹⁾ and, lastly, to maintain desirable characteristics in cattle⁽³⁾.

Embryo bisection has allowed to increase pregnancy rate⁽²⁹⁾ and the number of offspring⁽¹³⁾, compared to the transfer of whole embryos^(13,24). Most authors have reported a higher number of semi-embryos available for transfer in relation to the number of bisected embryos, thus increasing the efficiency in the number of offspring (Table 1). However, there is a great variation (75 to 118 %, efficiency), which could be mainly associated with factors related to the original embryo. In ewes, the pregnancy rate was 64 % when they received pairs of demi-embryos, obtaining 118 % efficiency in offspring⁽¹⁴⁾. Likewise, there was a higher percentage of embryo survival when transferring two demi-embryos per recipient ewe (101 %, 710/705), compared to the transfer of two whole embryos (62 %, 771/1252) considering the number of original embryos⁽²⁴⁾. Further, 30 % more lambs were born after the transfer of pairs of demi-embryos, compared with whole embryos (85 vs 55 %, $P<0.05$)⁽³⁰⁾.

Table 1: Efficiency of embryo bisection in the production of offspring in relation to the number of bisected embryos

Species	Bisected embryos	Number of offspring born	Efficiency (%)	Reference
Bovine	36	27	75	13
Bovine	50	61*	105	15
Bovine	11	12*	109	5
Sheep	40	34	85	30
Sheep	24	21**	88	16
Sheep	705	710*	101	24
Sheep	16	17	106	31
Sheep	39	46	118	14

* Number of fetuses diagnosed by ultrasound between 30 and 80 d of gestation or ** by sacrifice surgery after slaughter. Efficiency (%)= Number of offspring born / embryos bisected.

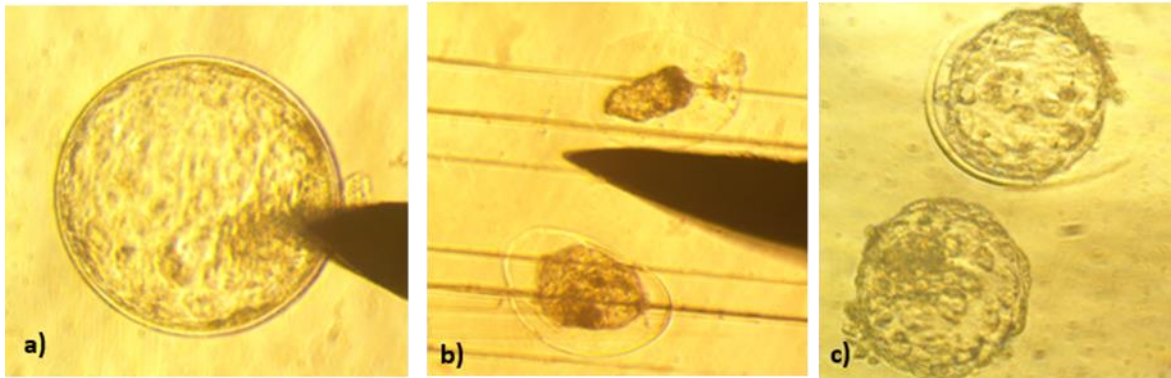
On the other hand, in some studies, low efficiency was obtained with embryo bisection^(16,32,33). It has been reported that a lower percentage of lambs were born after the transfer of bisected sheep embryos, compared to whole embryos (27 vs 52 %, $P<0.05$)⁽³⁴⁾. However, in beef cattle, despite the problems being associated with twin gestation⁽⁴⁾, implementing embryo bisection represents an economic benefit once the number of offspring is increased^(2,17,35). Therefore, embryo bisection can be implemented in embryo transfer programs^(2,24).

Embryonic reconstitution after bisection

Embryo reconstitution is an indicator of the ability of demi-embryos to become offspring after their transfer to a recipient female^(18,29). In adult tissues, stem cells are responsible for repairing injuries and regenerating tissues^(23,36) due to aging or diseases⁽³⁷⁾. In the case of embryos, something similar happens, through the replacement of specialized cells that have been lost due to some alteration⁽²³⁾. Embryos are capable of repairing their injuries, adapting to environmental conditions to survive after their reconstitution⁽²³⁾. Embryonic cells in early stages can adapt both in mitotic rate and differentiation process⁽³⁸⁾, due to the plasticity they present^(39,40). In addition, it has been proven that embryonic cell conglomerates in the early stages of development can become living organisms through cell reorganization, such as the case of blastocyst splitting⁽⁴¹⁾.

Thus, a group of cells has properties that exceed the potency of any of the individual cells within the group for cell reconstitution, which could be a joint effect⁽²³⁾. The extruded cells or even cell debris observed after embryo bisection could contain enough viable cells to proliferate and reorganize, originating another functional embryo⁽⁸⁾. At the time of embryo bisection, the resulting halves can be cultured *in vitro* for 2 to 48 h^(5,26). In each demi-embryo, the ICM is reorganized and the reconstitution of the blastocele begins immediately^(5,29,42). The union of the edges of trophoblastic cells during bisection is responsible for the trophoblast's ability to reconstitute itself⁽⁵⁾ since this group of cells secretes fluid into the blastocele, a process regulated by genes^(43,44). This allows the spherical formation of demi-embryos⁽⁴²⁾, being the trophoblastic cells important for embryo implantation⁽⁴⁵⁾. Likewise, an embryo, even if it has lost half of its cells, it is still an organism and a characteristic of organisms is to repair their lesions, regenerating them to continue their development⁽²³⁾. In the laboratory, it was bisected sheep expanded blastocysts produced *in vitro* with a microblade (Figure 1a), using the procedure called scratched bottom dish⁽⁴⁶⁾ (Figure 1b) and it was observed a completely spherical reconstitution (70 % the size of the original embryo) 12 h after *in vitro* culture (Figure 1c).

Figure 1: Embryo bisection process and reconstitution of the demi-embryos after 12 h of *in vitro* culture



a) Expanded blastocyst oriented symmetrically with respect to the bisecting microblade, b) resulting demi-embryos, and c) reconstituted demi-embryos. $\times 200$.

Several authors have reported a reconstitution percentage ranging from 90 to 178 % (Table 2).

Table 2: Reconstitution efficiency of bovine demi-embryos after *in vitro* culture for 2-48 h post-bisection

Bisected embryos	Number of demi-embryos	Reconstitution* (%)	Reference
21	19	90	29
11	16	145	5
176	268	152	12
19	30	158	47
230	408	178	28

*Reconstitution (%) = Number of demi-embryos / bisected embryos.

The evaluation of embryo reconstitution could allow the selection of viable demi-embryos and it can be a useful tool in embryo transfer programs⁽¹⁸⁾.

Factors that affect demi-embryo reconstitution

Embryo bisection technique

Embryo bisection is a technique that allows the production of identical twins in embryo transfer programs⁽⁵⁾. In the 80s, the technique required up to six embryo manipulation and

bisection instruments⁽⁵⁾. However, over time, different methodologies have been developed to simplify the technique⁽¹⁾, since the procedure required up to 15 min to bisect an embryo⁽⁴⁸⁾. In addition, the use of cutting instruments, such as microblade^(5,25,49) or glass needle^(50,51,52), have been extensively studied, with the objective of minimizing cellular damage at the time of cutting⁽⁵³⁾. Thus, the success of the technique depends on the minimum damage produced to the embryos⁽¹⁹⁾, since the procedure generates 10 to 13% of cell loss^(47,54).

On this regard, the microblade embryo bisection method has proven to be practical and with an application under field conditions⁽¹⁾. The implementation of this technique has been simplified by means of vertical pressure at the time of embryo bisection, using a microblade adapted to a single micromanipulator, without the use of an embryo holding micropipette^(12,15,34). In the laboratory, has been observed that the use of the scratched bottom dish technique⁽⁴⁶⁾ with 50 μ L of a commercial bisecting medium, facilitates embryonic fixation and prevents cell adhesion to bisecting and culture materials. This makes it possible to bisect groups of five embryos in approximately 3 min, making the application of this biotechnology more practical and without subjecting the embryos to prolonged stress.

On the other hand, there is evidence showing that the technician who performs the embryo bisection influences the productive response. In sheep, it has been evaluated the effect of the technician at the time of bisection, finding a significant difference between the two technicians in pregnancy rate (66 vs 75 %, $P<0.05$) and demi-embryo survival (51 vs 44 %, $P<0.01$)⁽⁵⁵⁾. Thus, proper training of the technician should be considered before implementing embryonic bisection.

Developmental stage

The stage of embryonic development, morula, early blastocyst, or expanded blastocyst at the time of bisection, is one of the most important factors that affect the pregnancy rate of transferred embryos⁽⁸⁾. After the bisection of mouse embryos, it was found that, in the morula stage, the demi-embryos are reconstituted in a lower percentage than in the blastocyst stage (74 vs 90 %, $P=0.001$) after *in vitro* culture for 24 h⁽⁴⁵⁾. In cattle, no significant differences were reported when transferring demi-embryos from morulae, early blastocysts, and expanded blastocysts, on pregnancy rate (51-65 %, $P>0.1$)⁽⁵³⁾. In another study, there was a lower pregnancy rate using bisected morulae (7/44, 16 %) compared to early blastocysts (58/96, 60 %), $P<0.01$ ⁽⁸⁾. Likewise, it has been reported a higher percentage of viable fetuses after blastocyst bisection compared to morulae, 91 (10/11) vs 30 % (3/10), $P<0.05$, at d 70 of gestation⁽⁴⁷⁾. Finally, in sheep, six pairs of identical twins were obtained from blastocyst bisection, without success in morula bisection ($P<0.05$)⁽¹⁶⁾.

In a practical way, it appears that bisecting morulae is easier due to the morphological symmetry they present, however, in the laboratory, it was observed that bisecting expanded blastocysts and even hatched blastocysts was easier once the ICM and TE were clearly identified. Furthermore, some authors have reported a higher pregnancy rate using blastocysts compared to morulae (Table 3), perhaps because they are more tolerant to manipulation and are less affected by the loss of the zona pellucida^(8,54). This could be due to the fact that embryogenesis is strictly regulated in time⁽²²⁾ and the more developed the embryos are, the more tolerant they are.

Table 3: Effect of the developmental stage of the whole embryo on the pregnancy rate of transferred semi-embryos

Species	Developmental stage, % (n)			Reference
	Morula	Blastocyst	Expanded blastocyst	
Caprine	0 (5)	33 (9)	55 (11)*	10
Ovine	60 (20)	88 (24)	-	16
Bovine	48 (162)	60 (96)	54 (28)	8
Bovine	51 (71)	64 (61)	58 (12)	53
Bovine	39 (139)	36 (33)	30 (10)	12

%= Pregnancy rate; n= Number of recipient females; *Hatched blastocyst.

Embryo quality

There is wide evidence for the use of excellent quality embryos for bisection purposes^(12,40,56,57). The embryos selected for bisection must meet certain morphological criteria, from which the success of demi-embryo reconstitution will depend⁽²⁸⁾ and, consequently, the pregnancy rate^(12,51). The quality of the embryos must be excellent or good, depending on the morphological criteria⁽⁵⁸⁾ because when they are of low quality (fair and bad) are more vulnerable to the bisection process^(12,47).

In bovines, when bisecting morulae, a higher percentage of survival was obtained in the group of excellent and good quality, compared to morulae of regular and poor quality, 167 (20/12) vs 75% (9/12), $P < 0.001$ ⁽⁴⁷⁾. On the other hand, 42 % pregnancy was found in cows after thawing and transferring excellent quality demi-embryos, while when transferring demi-embryos from low-quality embryos, no female became pregnant⁽⁵¹⁾. Likewise, it has been reported a higher percentage of development in pairs of demi-embryos, when bisecting bovine embryos of excellent quality, compared to those of good quality (76 vs 40 %, $P < 0.05$)⁽¹²⁾. Therefore, evaluation of embryo quality subjected to bisection should be

considered to obtain positive results. However, the morphological evaluation of embryos subjected to bisection is a subjective aspect, based on the experience of the researcher.

In the experience in manipulating sheep embryos of this research team, was found a higher percentage of demi-embryo reconstitution (145 %, approximately), when bisecting embryos of excellent quality and with a diameter greater than 230 μm . In the resulting demi-embryos, after 12 h of *in vitro* culture, was found that when they were reconstituted, they had an average diameter of $176 \pm 10.03 \mu\text{m}$ (Figure 2), similar to that reported in quality two porcine demi-embryos ($161.6 \pm 25.7 \mu\text{m}$)⁽²⁶⁾, but higher than that reported in human demi-embryos ($121 \mu\text{m}$)⁽²²⁾. Therefore, the embryonic diameter has been proposed as an indicator of quality^(25,59) since the size of the embryo is important in maternal recognition⁽⁶⁰⁾.

On the other hand, the size of the embryo is also associated with the number of cells⁽⁶¹⁾ and, consequently, with the embryo quality. In poor quality embryos, there is a deficiency in the rate of cell division, mitosis, and consequently in the number of cells⁽⁴⁷⁾. Thus, the size of the embryo is proportional to the number of cells used for its reconstitution⁽²²⁾. There is a 50 % recovery of cells from the original embryos in the resulting demi-embryos depending on the quality and uniformity of the bisection process^(26,28). This suggests that bisecting bigger size embryos will produce semi-embryos with more ICM and TE cells, thus increasing their survival. In the laboratory, was obtained an average of 68 ± 11.3 cells in reconstituted demi-embryos, after 12 h of *in vitro* culture (Figure 3), from embryos with 122 ± 6.6 cells. In this sense, active cell proliferation can be a criterion of embryonic quality⁽⁶²⁾. Based on the above, the diameter of the embryo could be an objective criterion to select embryos for bisection, in order to achieve the greatest possible success in demi-embryos reconstitution.

Figure 2. *In vitro* reconstitution of demi-embryos after 12 h of culture. $\times 200$

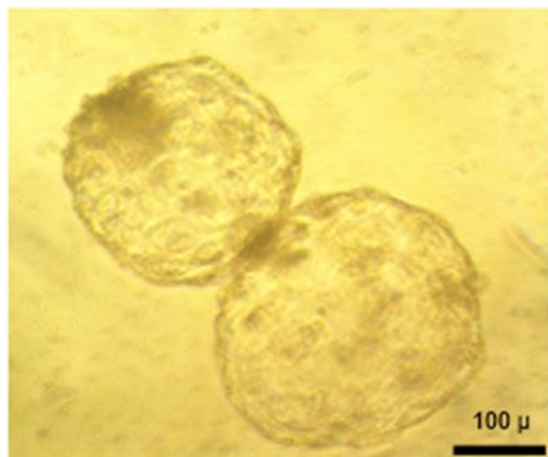
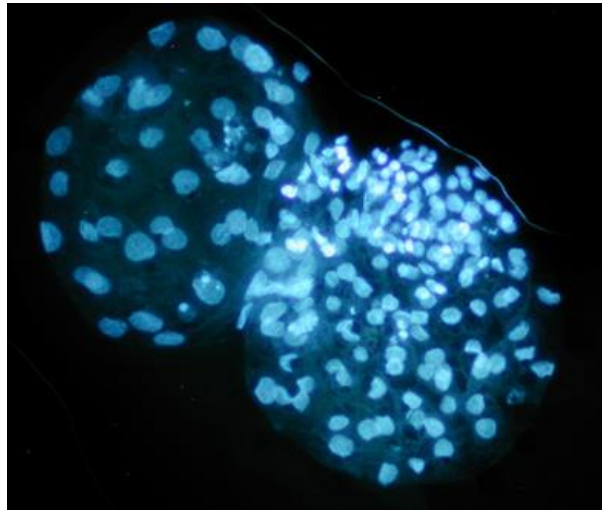


Figure 3. Cell staining (Hoechst) of *in vitro* demi-embryos after 12 h of culture. $\times 200$ 

***In vitro* or *in vivo* produced embryos**

There are differences between embryos produced *in vitro* or *in vivo*, both in morphology and in molecular components⁽⁶³⁾, where *in vivo* produced embryos are of better quality. Nevertheless, a high percentage of *in vitro* survival of ovine demi-embryos has been reported after bisection (80-85 %), and 33 % pregnancy after transferring pairs of demi-embryos to recipient ewes (5/15)⁽³⁴⁾. On the other hand, for embryos produced *in vivo*, some authors reported greater pregnancy rates. In bisected bovine embryos produced *in vivo* and cultured the demi-embryos *in vitro*, a high percentage of reconstitution was reported (47/60, 78.3 %)⁽¹⁵⁾. In bovines, there was 53 % (18/34) pregnancy when transferring embryos produced *in vivo*, bisected, and cultured *in vitro*⁽¹⁸⁾. Thus, the efficiency of embryo bisection in relation to the origin of the embryo seems to be lower in embryos from *in vitro* conditions. This could be due to the low quality and efficiency of *in vitro* production⁽⁶³⁾. Therefore, it is necessary to improve efficiency in both embryo production procedures, once the two of them are focused on improving productivity in livestock production⁽⁶⁴⁾.

Effect of breed and age of embryo donors for bisection

There are other little-studied factors that could affect the embryo's fate under the bisection process. In sheep, the effect of the breed was evaluated, without finding significant differences in pregnancy rate and survival of demi-embryos between embryos from Gotland

and Finnish Texel (69 vs 50 % and 42 vs 26 %, respectively, $P>0.05$) and between Danish Texel and Finnish Texel breeds (74 vs 74 % and 50 vs 50 %, respectively, $P>0.05$)⁽⁵⁵⁾. In addition, the effect of the donor age has been reported, without finding differences in pregnancy rate between embryos generated from adult females (24 months of age) and young (approximately 10 mo of age) (74 vs 74 %, $P>0.05$), however, demi-embryo survival (51 vs 47, $P<0.05$) and percentage of identical twins was higher in adult ewes than in young (38 vs 27, $P<0.01$)⁽⁵⁵⁾. This could be related to a lower survival capacity of whole embryos from young ewes^(65,66,67), which is confirmed after transferring demi-embryos⁽⁵⁵⁾.

Conclusions

Demi-embryos reconstitution is a key factor for the success of embryo bisection and the highest efficiency is obtained by selecting excellent quality embryos regardless of their developmental stage. The results of the literature show the potential of the bisection technique; therefore, its application should be considered to improve the efficiency of embryo transfer programs.

Literature cited:

1. Godke RA, Sansinena M, Youngs CR. Assisted reproductive technologies and embryo culture methods for farm animals. In: Pinkert CA editor. Transgenic Animal Technology: A Laboratory handbook. 3rd ed. Amsterdam: Elsevier; 2014:581-638.
2. Praharani L. Factors affecting twinning and the impacts of twinning in cattle. WARTAZOA Indones Bull Anim Vet Sci 2019;29(1):13-24.
3. Yang X, Tian XC, Kubota C, Page R, Xu J, Cibelli J, *et al*. Risk assessment of meat and milk from cloned animals. Nat Biotechnol 2007;25(1):77-83.
4. Wakchaure R, Ganguly S. Twinning in cattle: A Review. ARC J Gynecol Obstet 2016;1(4):1-3.
5. Ozil JP. Production of identical twins by bisection of blastocysts in the cow. Reproduction 1983;69(2):463-468.
6. Escriba MJ, Valbuena D, Remohí J, Pellicer A, Simon C. New techniques on embryo manipulation. J Reprod Immunol 2002;55(1-2):149-161.
7. Tang HH, Tsai YC, Kuo CT. Embryo splitting can increase the quantity but not the quality of blastocysts. Taiwan J Obstet Gynecol 2012;51(2):236-239.

8. Williams TJ, Elsdon RP, Seidel Jr GE. Pregnancy rates with bisected bovine embryos. *Theriogenology* 1984;22(5):521-531.
9. Illmensee K, Levanduski M. Embryo splitting. *Middle East Fertil Soc J* 2010;15(2):57-63.
10. Tsunoda Y, Tokunaga T, Sugie T, Katsumata M. Production of monozygotic twins following the transfer of bisected embryos in the goats. *Theriogenology* 1985;24(3):337-343.
11. Ramón UJ, Meza VV, Deneb CP, Domínguez RA, Quintal FJ. Bisection and embryo transfer in hair sheep. *Biotechnology Summit 2012, Mérida, Yucatán, México 2012*;12-21(3):138-142.
12. Hashiyada Y. The contribution of efficient production of monozygotic twins to beef cattle breeding. *J Reprod Develop* 2017;63(6):527-538.
13. Dahlen CR, DiCostanzo A, Spell AR, Lamb GC. Use of embryo transfer seven days after artificial insemination or transferring identical demi-embryos to increase twinning in beef cattle. *J Anim Sci* 2012;90(13):4823-4832.
14. Chesne P, Colas G, Cognie Y, Guerin Y, Sévellec C. Lamb production using superovulation, embryo bisection, and transfer. *Theriogenology* 1987;27(5):751-757.
15. Saito S, Niemann H. *In vitro* and *in vivo* survival of bovine demi-embryos following simplified bisection and transfer of one or two halves per recipient. *J Reprod Develop* 1993;39(3):251-258.
16. Shelton JN. Factors affecting viability of fresh and frozen-thawed sheep demi-embryos. *Theriogenology* 1992;37(3):713-721.
17. Echtenkamp SE, Gregory KE. Reproductive, growth, feedlot, and carcass traits of twin vs single births in cattle. *J Anim Sci* 2002;80:E64-E73.
18. Alvarez RH, Pires RML, Campanha A, Oba E. Short-term culture of bovine bisected embryos. Effects on pregnancy rates, sex ratio and birth weight of calves. *B Indústr Anim* 2008;65(3):191-196.
19. Skrzyszowska M, Smorag Z, Katska L. Demi-embryo production from hatching of zona-drilled bovine and rabbit blastocysts. *Theriogenology* 1997;48(4):551-557.
20. Reichenbach HD, Schwartz J, Wolf E, Brem G. Effects of embryo developmental stage, quality and short-term culture on the efficiency of bovine embryo splitting. *Theriogenology* 1998;1(49):224.

21. Kawachiya S, Bodri D, Shimada N, Kato K, Takehara Y, Kato O. Blastocyst culture is associated with an elevated incidence of monozygotic twinning after single embryo transfer. *Fertil Steril* 2011;95(6):2140-2142.
22. Noli L, Ogilvie C, Khalaf Y, Ilic D. Potential of human twin embryos generated by embryo splitting in assisted reproduction and research. *Hum Reprod* 2017;23(2):156-165.
23. Condic ML. Totipotency: what it is and what it is not. *Stem Cells Dev* 2014;23(8):796-812.
24. Vivanco HW, Rangel SR, Lynch P, Rhodes A. Large scale commercial application of bisection of sheep embryos. *Theriogenology* 1991;35(1):292.
25. McKinnon AO, Carnevale EM, Squires EL, Carney NJ, Seidel Jr GE. Bisection of equine embryos. *Equine Vet J* 1989;21(Suppl 8):129-133.
26. Reichelt B, Niemann H. Generation of identical twin piglets following bisection of embryos at the morula and blastocyst stage. *Reproduction* 1994;100(1):163-172.
27. Yang X, Anderson GB. Micromanipulation of mammalian embryos: principles, progress and future possibilities. *Theriogenology* 1992;38(2):315-335.
28. Rho GJ, Johnson WH, Betteridge KJ. Cellular composition and viability of demi-and quarter-embryos made from bisected bovine morulae and blastocysts produced *in vitro*. *Theriogenology* 1998;50(6):885-895.
29. Baker RD, Shea BF. Commercial splitting of bovine embryos. *Theriogenology* 1985;23(1):3-12.
30. Vintila I, Bencsik I, Pacala N, Corin N, Babusik I, Kulickova L. Embryo splitting- a way to increase the efficiency of embryo-transfer in sheep. *Stočarstvo: Časopis za unapređenje stočarstva* 1995;49(9-12):349-353.
31. Széll A, Hudson RHH. Factors affecting the survival of bisected sheep embryos *in vivo*. *Theriogenology* 1991;36(3):379-387.
32. Shelton JN, Szell A. Survival of sheep demi-embryos *in vivo* and *in vitro*. *Theriogenology* 1988;30(5):855-863.
33. Harkness UF, Crombleholme TM. Twin–twin transfusion syndrome: where do we go from here? *Semin Perinato* 2005;29(5):296-304.

34. Morton KM, Rowe AM, Maxwell WC, Evans G. *In vitro* and *in vivo* survival of bisected sheep embryos derived from frozen-thawed unsorted, and frozen-thawed sex-sorted and refrozen-thawed ram spermatozoa. *Theriogenology* 2006;65(7):1333-1345.
35. De Rose EP, Wilton JW. Productivity and profitability of twin births in beef cattle. *J Anim Sci* 1991;69(8):3085-3093.
36. Kenyon J, Gerson SL. The role of DNA damage repair in aging of adult stem cells. *Nucleic Acids Res* 2007;35(22):7557-7565.
37. Maynard S, Fang EF, Scheibye-Knudsen M, Croteau DL, Bohr VA. DNA damage, DNA repair, aging, and neurodegeneration. *Cold Spring Harb Perspect Med* 2015;5(10):a025130.
38. Deuchar EM. Regeneration of amputated limb-buds in early rat embryos. *Development* 1976;35(2):345-354.
39. Cenariu M, Pall E, Cernea C, Groza I. Evaluation of bovine embryo biopsy techniques according to their ability to preserve embryo viability. *J Biomed Biotechnol* 2012;2012.
40. De Sousa RV, da Silva Cardoso CR, Butzke G, Dode MAN, Rumpf R, Franco MM. Biopsy of bovine embryos produced *in vivo* and *in vitro* does not affect pregnancy rates. *Theriogenology* 2017;90:25-31.
41. Mitalipov S, Wolf D. Totipotency, pluripotency and nuclear reprogramming. In: Martin U. editor. *Engineering of stem cells*. Heidelberg, Berlín, Alemania: Springer; 2009;114:185-199.
42. Daniel Jr JC. Some kinetics of blastocyst formation as studied by the process of reconstitution. *J Exp Zool* 1963;154(2):231-237.
43. Watson AJ, Barcroft LC. Regulation of blastocyst formation. *Front Biosci* 2001;6:D708-D730.
44. Watson AJ, Natale DR, Barcroft LC. Molecular regulation of blastocyst formation. *Anim Reprod Sci* 2004;82:583-592.
45. Wang ZJ, Trounson A, Dziadek M. Developmental capacity of mechanically bisected mouse morulae and blastocysts. *Reprod Fertil Dev* 1990;2(6):683-691.
46. Bredbacka P. Biopsy of morulae and blastocysts. *Reprod Domest Anim* 1991;26(2):82-84.

47. McEvoy TG, Sreenan JM. Effect of embryo quality and stage of development on the survival of zona pellucida-free cattle demi-embryos. *Theriogenology* 1990;33(6):1245-1253.
48. Yang X, Foote RH. Production of identical twin rabbits by micromanipulation of embryos. *Biol Reprod* 1987;37(4):1007-1014.
49. Lopes RFF, Forell F, Oliveira ATD, Rodrigues JL. Splitting and biopsy for bovine embryo sexing under field conditions. *Theriogenology* 2001;56(9):1383-1392.
50. Willadsen SM, Godke RA. A simple procedure for the production of identical sheep twins. *Vet Rec* 1984;114(10):240-243.
51. Niemann H, Brem G, Sacher B, Smidt D, Kräusslich H. An approach to successful freezing of demi-embryos derived from day-7 bovine embryos. *Theriogenology* 1986;25(4):519-524.
52. Seike N, Saeki K, Utaka K, Sakai M, Takakura R, Nagao Y, *et al.* Production of bovine identical twins via transfer of demi-embryos without zonae pellucidae. *Theriogenology* 1989;32(2):211-220.
53. Kippax IS, Christie WB, Rowan TG. Effects of method of splitting, stage of development and presence or absence of zone pellucida on foetal survival in commercial bovine embryo transfer of bisected embryos. *Theriogenology* 1991;35(1):25-35.
54. Skrzyszowska M, Smorag Z. Cell loss in bisected mouse, sheep and cow embryos. *Theriogenology* 1989;32(1):115-122.
55. Rangel-Santos R. Investigations into procedures for the implementation of a multiple ovulation and embryo transfer scheme using ewe lambs [PhD thesis]. Wellington, New Zealand: Massey University; 1991.
56. Shea BF. Determining the sex of bovine embryos using polymerase chain reaction results: a six-year retrospective study. *Theriogenology* 1999;51(4):841-854.
57. Lopatarova M, Cech S, Krontorad P, Holy L, Hlavicova J, Dolezel R. Sex determination in bisected bovine embryos and conception rate after the transfer of female demi-embryos. *Vet Med* 2008;53(11):595-603.
58. Stringfellow DA, Seidel G. *Manual of the International Embryo Transfer Society*. 2nd. IETS 1990;19.
59. Mori M, Otoi T, Suzuki T. Correlation between the cell number and diameter in bovine embryos produced *in vitro*. *Reprod Domest Anim* 2002;37(3):181-184.

60. Goff AK. Embryonic signals and survival. *Reprod Domest Anim* 2002;37(3):133-139.
61. O'Hara L, Forde N, Kelly AK, Lonergan P. Effect of bovine blastocyst size at embryo transfer on Day 7 on conceptus length on Day 14: can supplementary progesterone rescue small embryos? *Theriogenology* 2014;81(8):1123-1128.
62. Makarevich AV, Markkula M. Apoptosis and cell proliferation potential of bovine embryos stimulated with insulin-like growth factor I during *in vitro* maturation and culture. *Biol Reprod* 2002;66(2):386-392.
63. Camargo LSDA, Viana JHM, Sá WFD, Ferreira ADM, Ramos ADA, Vale Filho VR. Factors influencing *in vitro* embryo production. *Anim Reprod* 2006;3(1):19-28.
64. Paramio MT. *In vivo* and *in vitro* embryo production in goats. *Small Ruminant Res* 2010;89(2-3):144-148.
65. Quirke JF, Hanrahan JP. Comparison of the survival in the uteri of adult ewes of cleaved ova from adult ewes and ewe lambs. *Reproduction* 1977;51(2):487-489.
66. McMillan WH, McDonald MF. Survival of fertilized ova from ewe lambs and adult ewes in the uteri of ewe lambs. *Anim Reprod Sci* 1985;8(3):235-240.
67. Morton KM. Developmental capabilities of embryos produced *in vitro* from prepubertal lamb oocytes. *Reprod Domest Anim* 2008;43(Suppl 2):137-143.